



ICSI with testicular sperm for couples with sperm DNA damage

Armand Zini¹

¹ *Department of Surgery, St. Mary's Hospital, McGill University, Montreal, Canada*

The advent of intracytoplasmic sperm injection (ICSI) has revolutionized the management of male factor infertility (1). Shortly after the technique was introduced, studies demonstrated that ICSI could successfully treat couples with severe male factor infertility. Several investigators reported that neither sperm concentration, morphology, nor progressive motility had any impact on ICSI outcomes (2-4). However, the only sperm characteristic that portended a negative ICSI outcome was the injection of a totally immotile (and presumably dead) spermatozoon (4-6).

More recent publications have shown that ICSI may not overcome significant sperm abnormalities. Mitchell et al. reported significantly lower clinical pregnancy rates in couples with sperm motility <5% compared to those with higher sperm motility (11% vs. 41%, $P=0.04$ for couples with sperm motility <5% and >5%, respectively) (7). De Vos et al. reported significantly higher clinical pregnancy (37% vs. 20%, respectively, $P=0.018$), implantation (32% vs. 23%, respectively, $P=0.013$) and live birth rates (28% vs. 20%, respectively, $P=0.006$) with the use of morphologically normal vs. morphologically abnormal sperm for ICSI (8). Strassburger et al. studied 1,076 unselected ICSI cycles and reported that cryptozoospermic couples had significantly lower fertilization and clinical pregnancy rates (46% vs. 61%, $P<0.0001$ and 20% vs. 31%, $P<0.05$, respectively) and higher miscarriage rates (30% vs. 15%, $P<0.03$) when compared to couples with a sperm concentration between 1×10^5 sperm/mL and 1×10^7 sperm/mL (9).

Subfertile men with abnormal semen parameters may have an underlying sperm genetic defect that could potentially impact on IVF and ICSI outcomes. Indeed, studies have shown that sperm DNA damage is more common in men with poor semen parameters than in those with normal parameters and in 2008 a systematic review and meta-analysis of 2,162 IVF and ICSI treatment cycles demonstrated a potential adverse effect of sperm DNA damage on the chance of pregnancy, with a diagnostic odds ratio of 1.44 (95% CI, 1.03, 2.03) (10, 11). Moreover, a study of 1,549 IVF/ICSI cycles concluded that sperm DNA damage was predictive of pregnancy loss after IVF/ICSI (combined OR 2.48; 95% CI 1.52-4.04; $P<0.0001$) (12). More recent meta-analyses have similarly reported that sperm DNA damage is associated with lower clinical pregnancy rates and higher miscarriage rates after IVF and ICSI (13-15).

In 2005, Greco et al., reported their experience with the use of testicular sperm with ICSI in a small

series of couples with sperm DNA damage (16). Greco et al., claimed that there was “no specific treatment” for sperm DNA damage and hypothesized that the DNA damage in ejaculated sperm begins after spermatozoa are released from Sertoli cells. In view of the adverse impact of sperm DNA damage on IVF and ICSI outcomes, Greco et al. proposed using testicular sperm in men with sperm DNA damage with the idea that sperm recovered directly from the testis would show less damage than ejaculated sperm. To test this idea or hypothesis, they (1) compared the DNA damage in ejaculated and testicular sperm in two sequential assisted reproduction cycles performed in couples with high levels of ejaculated sperm DNA damage and (2) evaluated the ICSI outcomes of these couples. Greco et al. reported higher pregnancy rates with ICSI when using testicular rather than ejaculated sperm in couples with sperm DNA damage and observed a higher frequency of sperm showing detectable DNA damage in ejaculated vs. testicular sperm (16). They suggested that the poorer outcome with ejaculated sperm was a result of acquired DNA damage during transit through the epididymis or possibly during ejaculation.

In 2005, Suganuma et al. conducted experimental studies using an animal model with abnormal spermatogenesis (mutant mice with minimal levels of transition nuclear proteins and incomplete sperm nuclear compaction). They planned to investigate whether DNA damage in ejaculated sperm is increased after spermatozoa are released from the testis (17). Suganuma et al. observed that in animals with abnormal spermatogenesis the passage of sperm through the epididymis was associated with a loss of sperm DNA integrity and fertilizing capacity (17). They speculated that in animals with poor sperm nuclear compaction, the sperm DNA is not fully protected during epididymal passage. In contrast, in animals with normal spermatogenesis, the passage of sperm through the epididymis was not associated with a similar loss of sperm DNA integrity and fertilizing capacity. As such, Suganuma et al. proposed that in some men (i.e. those with defective spermatogenesis) the passage of sperm through the epididymis could result in a loss of sperm DNA integrity and fertilizing capacity. This

concept (adverse effect of epididymal transit on sperm nuclear integrity) is contrary to the understanding of the biology of sperm maturation, specifically, the acquisition of sperm motility and nuclear compaction during passage of spermatozoa through the epididymis (18, 19).

The idea that the post-testicular environment or epididymal transit can induce sperm damage has led clinicians to utilize testicular rather than ejaculated sperm for ICSI in men with abnormal spermatogenesis and poor sperm DNA integrity. In a recent online survey of Canadian fertility clinics, 70% of the respondents reported performing testicular sperm retrieval (TSR) with ICSI for non-azoospermic men with poor sperm DNA integrity (Zini et al., unpublished observations). Similarly, over 70% of the respondents attending a session on testicular sperm for ICSI at the 2017 annual meeting of the American Society for Reproductive Medicine reported that they would (in selected cases) opt for testicular sperm rather than ejaculated sperm ICSI in men with sperm DNA damage (Zini et al., unpublished observations). However, despite the widespread utilization of TSR with ICSI in these men, there is no consensus or guideline on how to manage these cases in clinical practice.

In this issue of the International Brazilian Journal of Urology, Dr. Sandro Esteves and Dr. Mark Sigman present the Pro and Con perspectives, respectively, of using testicular rather than ejaculated sperm in non-azoospermic couples with high sperm DNA fragmentation (20, 21). Dr. Esteves discusses the adverse impact of sperm DNA damage on reproductive outcomes and demonstrates that sperm DNA damage is lower in testicular than in ejaculated sperm. Dr. Esteves presents a systematic of the literature and provides us with compelling evidence in support of the use of testicular rather than ejaculated sperm in these couples. Dr. Sigman argues that the quality of the studies on testicular sperm-ICSI is moderate at best and that there is insufficient evidence to adopt the practice of testicular sperm retrieval in these couples. Moreover, Dr. Sigman also cautions that sperm DNA testing requires further validation as a diagnostic test and in this context as well.

REFERENCES

1. Palermo G, Joris H, Devroey P, Van Steirteghem AC. Pregnancies after intracytoplasmic injection of single spermatozoon into an oocyte. *Lancet*. 1992;340(8810):17-8.
2. Palermo G, Joris H, Derde MP, Camus M, Devroey P, Van Steirteghem A. Sperm characteristics and outcome of human assisted fertilization by subzonal insemination and intracytoplasmic sperm injection. *Fertil Steril*. 1993;59:826-35.
3. Mansour RT, Aboulghar MA, Serour GI, Amin YM, Ramzi AM. The effect of sperm parameters on the outcome of intracytoplasmic sperm injection. *Fertil Steril*. 1995;64:982-6.
4. Nagy ZP, Liu J, Joris H, Verheyen G, Tournaye H, Camus M, et al. The result of intracytoplasmic sperm injection is not related to any of the three basic sperm parameters. *Hum Reprod*. 1995;10:1123-9.
5. Nijs M, Vanderzwalmen P, Vandamme B, Segal-Bertin G, Lejeune B, Segal L, et al. Fertilizing ability of immotile spermatozoa after intracytoplasmic sperm injection. *Hum Reprod*. 1996;11:2180-5.
6. Vandervorst M, Tournaye H, Camus M, Nagy ZP, Van Steirteghem A, Devroey P. Patients with absolutely immotile spermatozoa and intracytoplasmic sperm injection. *Hum Reprod*. 1997;12:2429-33.
7. Mitchell V, Rives N, Albert M, Peers MC, Selva J, Clavier B, et al. Outcome of ICSI with ejaculated spermatozoa in a series of men with distinct ultrastructural flagellar abnormalities. *Hum Reprod*. 2006;21:2065-74.
8. De Vos A, Van De Velde H, Joris H, Verheyen G, Devroey P, Van Steirteghem A. Influence of individual sperm morphology on fertilization, embryo morphology, and pregnancy outcome of intracytoplasmic sperm injection. *Fertil Steril*. 2003;79:42-8.
9. Strassburger D, Friedler S, Raziel A, Schachter M, Kasterstein E, Ron-el R. Very low sperm count affects the result of intracytoplasmic sperm injection. *J Assist Reprod Genet*. 2000;17:431-6.
10. Collins JA, Barnhart KT, Schlegel PN. Do sperm DNA integrity tests predict pregnancy with in vitro fertilization? *Fertil Steril*. 2008;89:823-31.
11. Zini A, Sigman M. Are tests of sperm DNA damage clinically useful? Pros and cons. *J Androl*. 2009;30:219-29.
12. Zini A, Boman JM, Belzile E, Ciampi A. Sperm DNA damage is associated with an increased risk of pregnancy loss after IVF and ICSI: systematic review and meta-analysis. *Hum Reprod*. 2008;23:2663-8.
13. Zhao J, Zhang Q, Wang Y, Li Y. Whether sperm deoxyribonucleic acid fragmentation has an effect on pregnancy and miscarriage after in vitro fertilization/ intracytoplasmic sperm injection: a systematic review and meta-analysis. *Fertil Steril*. 2014;102:998-1005.e8.
14. Oleszczuk K, Giwercman A, Bungum M. Sperm chromatin structure assay in prediction of in vitro fertilization outcome. *Andrology*. 2016;4:290-6.
15. Simon L, Zini A, Dyachenko A, Ciampi A, Carrell DT. A systematic review and meta-analysis to determine the effect of sperm DNA damage on in vitro fertilization and intracytoplasmic sperm injection outcome. *Asian J Androl*. 2017;19:80-90.
16. Greco E, Scarselli F, Iacobelli M, Rienzi L, Ubaldi F, Ferrero S, et al. Efficient treatment of infertility due to sperm DNA damage by ICSI with testicular spermatozoa. *Hum Reprod*. 2005;20:226-30.
17. Suganuma R, Yanagimachi R, Meistrich ML. Decline in fertility of mouse sperm with abnormal chromatin during epididymal passage as revealed by ICSI. *Hum Reprod*. 2005;20:3101-8.
18. Cooper TG. Sperm maturation in the epididymis: a new look at an old problem. *Asian J Androl*. 2007;9:533-9.
19. Sullivan R, Mieusset R. The human epididymis: its function in sperm maturation. *Hum Reprod Update*. 2016;22:574-87.
20. Esteves SC. Testicular versus ejaculated sperm should be used for intracytoplasmic sperm injection (ICSI) in cases of infertility associated with sperm DNA fragmentation | Opinion: Yes. *Int Braz J Urol*. 2018;44:667-75.
21. Sigman M. Testicular versus ejaculated sperm should be used for intracytoplasmic sperm injection (ICSI) in cases of infertility associated with sperm DNA fragmentation | Opinion: No. *Int Braz J Urol*. 2018;44:676-9.

Armand Zini, MD

Department of Surgery,
St. Mary's Hospital,
McGill University, Montreal, Canada
E-mail: ziniarmand@yahoo.com